



Université d'Orléans

Institut de Chimie Organique et Analytique

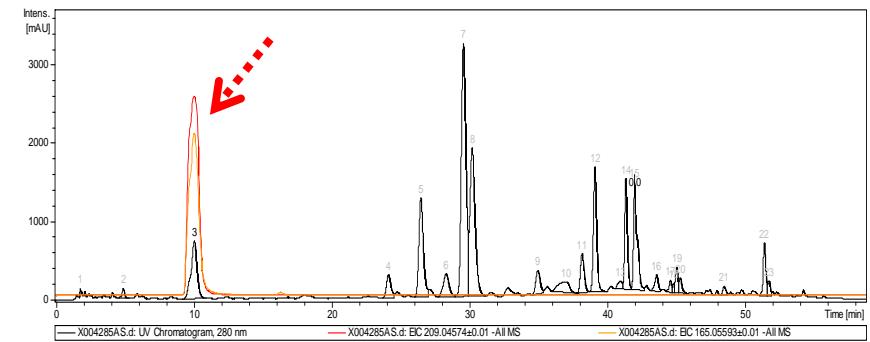
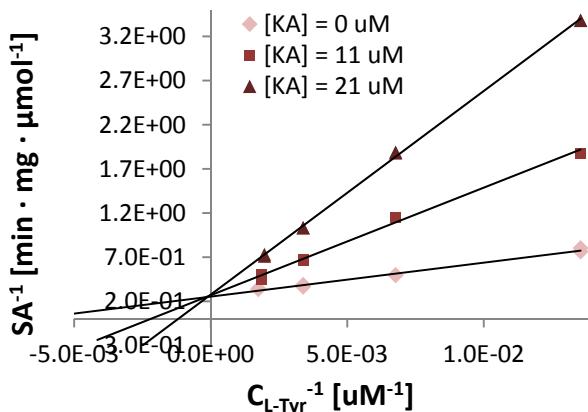
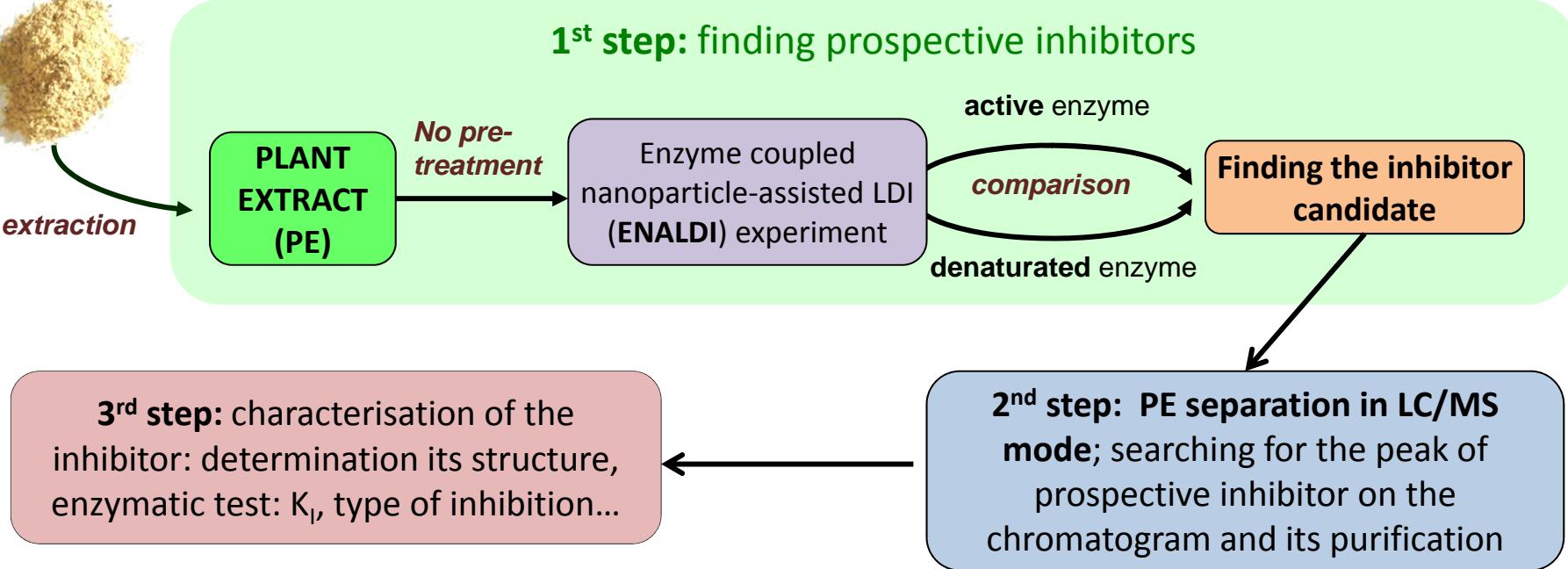
'Intensity ion fading-SALDI MS approach for searching inhibitors of tyrosinase in complex mixtures'

Aleksander Salwiński

Prof. Benoit Maunit, Prof. Raphaël Delépée



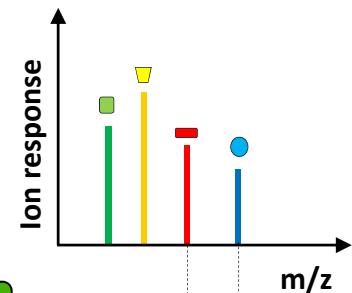
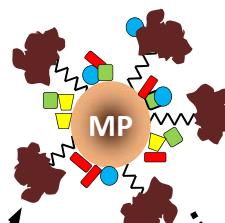
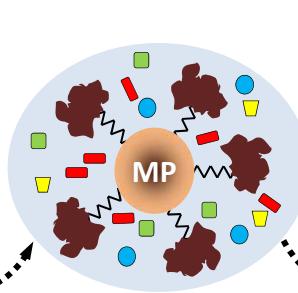
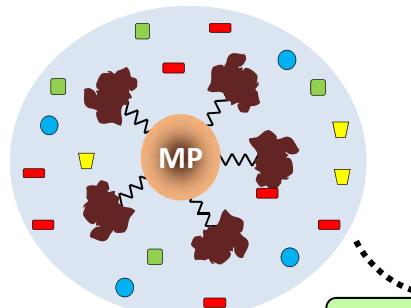
General outlook of the new approach



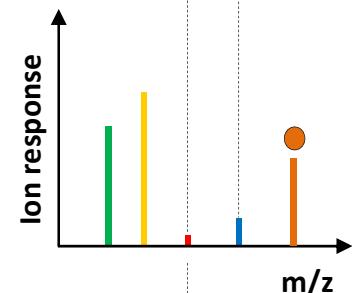
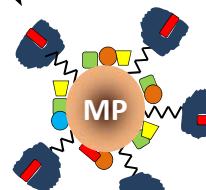
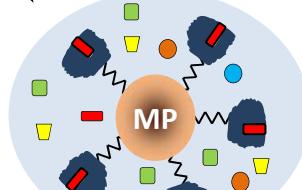
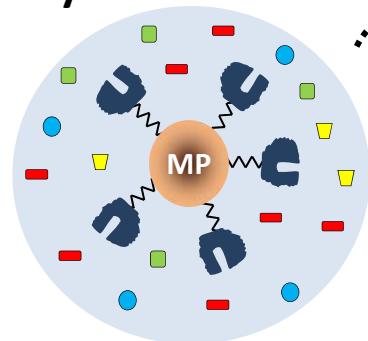
Enzyme coupled nanoparticle-assisted LDI (ENALDI) principle



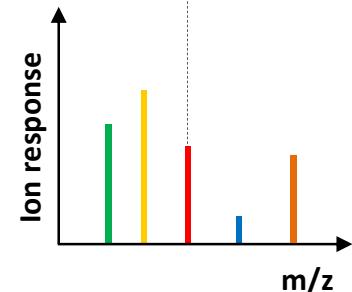
Denatured enzyme (control)



Active enzyme

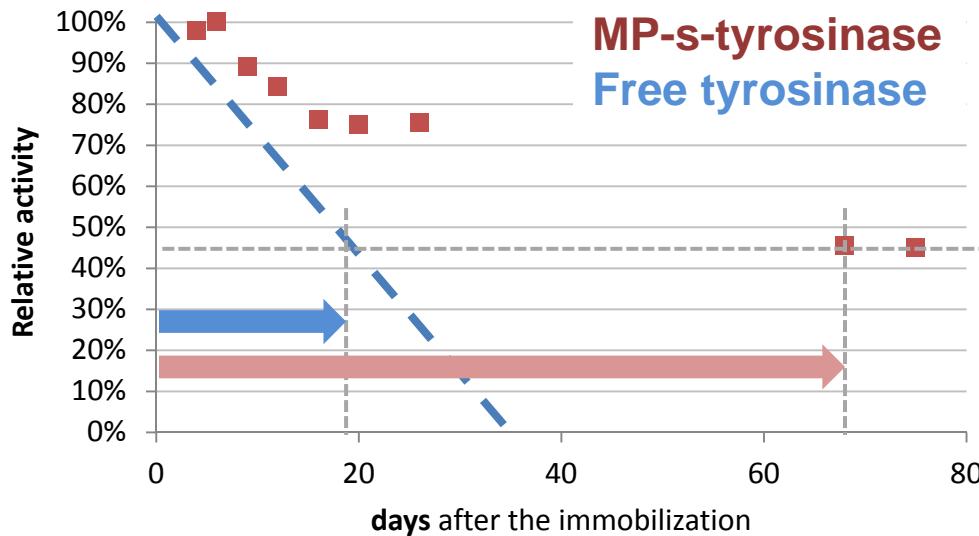


*denaturation on the MALDI spot
after drying by deposition 0.1% FA =
release of previously bound ligand*



MPs characterisation

- **Stability:**



Free tyrosinase loses **55% of the initial activity** within approx. 17-20 days [1,2] when stored at 4°C. MPs-immobilized tyrosinase loses 55% of initial activity up to 70-75 days.

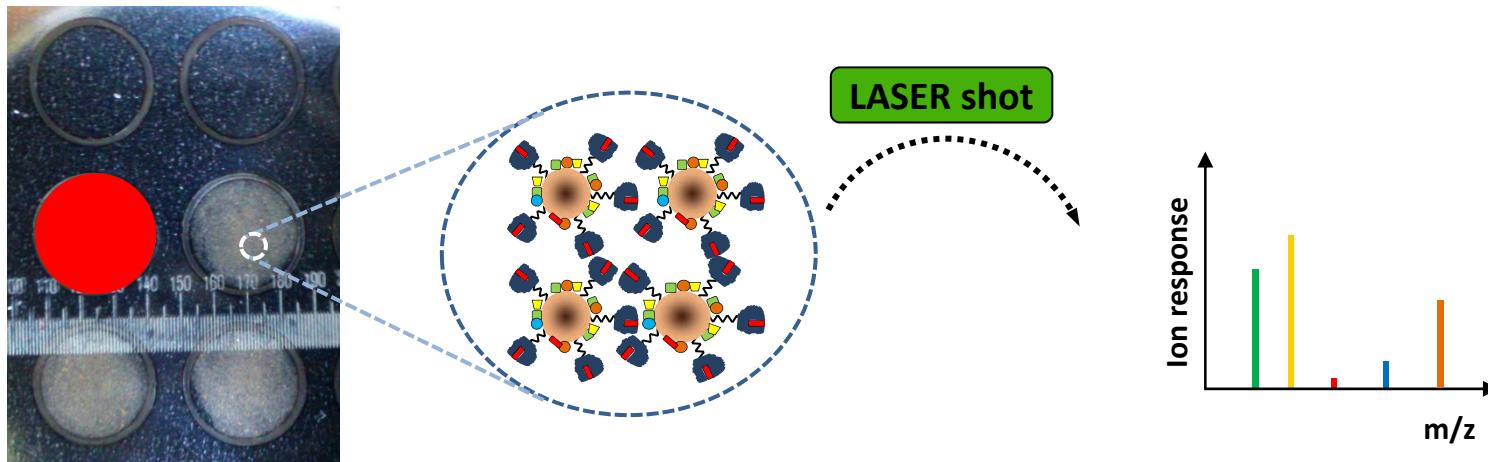
- **Protein binding capacity:**

245 mg of BSA/g of MPs

[1] A.M. Girelli, E. Mattei, A. Messina, D. Papaleo, *Sensors and Actuators B*, **2007**, 125, 48–54

[2] M. Y. Arıca, G. Bayramoglu, N. Bıçak, *Process Biochemistry*, **2004**, 39, 2007–2017

Enzyme coupled nanoparticle-assisted LDI (ENALDI) highlights



On a single spot:

- Total amount of MPs: **3.47 µg**;
- amount of tyrosinase enzyme bound to MPs: approx. **800 ng (6.5 pmol)**;
- amount of plant extract (equivalent of dry mass of plant powder): **300-450 ng**;

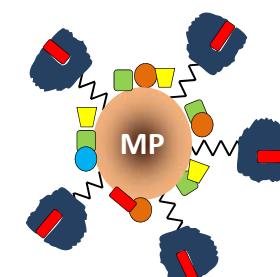
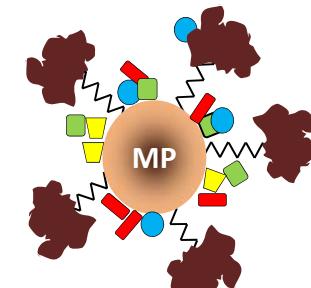
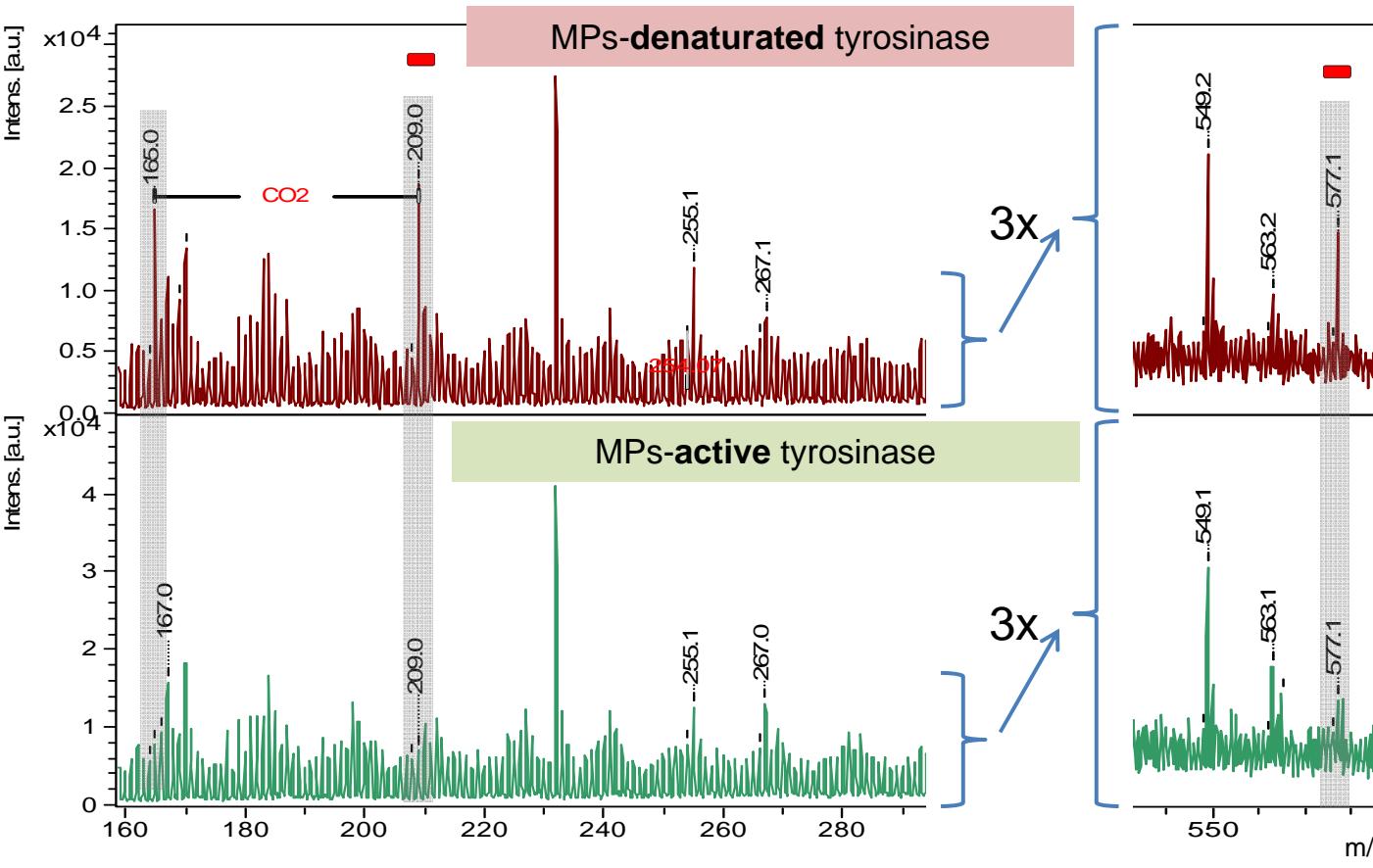
Dual function of magnetic nanoparticles:

- enzyme carrier;
- energy absorber: nanoparticle-assisted ionization is believed to be generated by heat released upon the laser shot

Experimental:

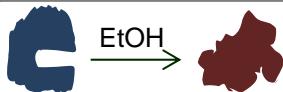
- Bruker Autoflex with MTP 384 standard target plate (polished steel)
- number of LASER shots: 300
- MS spectra collected in a negative mode

Exemplary ENALDI spectra of raw aqueous extract of licorice (*Glycyrrhiza glabra*) root



- significant drop of the ion response for 209.0 and 577.1 m/z ions

Enzyme coupled nanoparticle-assisted LDI (ENALDI) principle



Tyrosinase
active/denatured

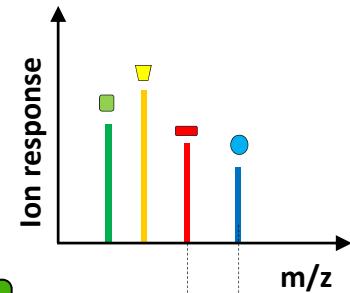
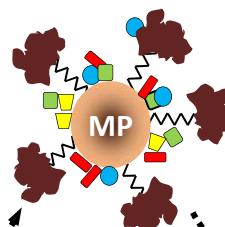
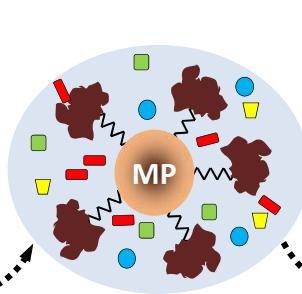
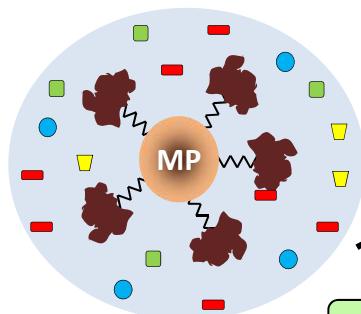
non-interacting
compounds

substrate

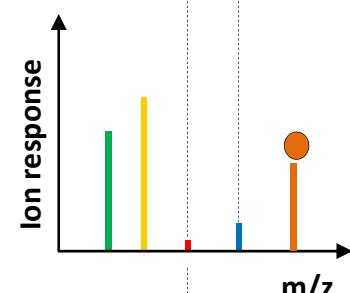
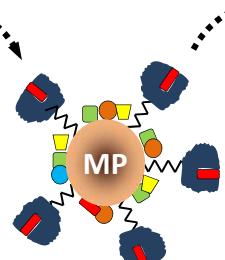
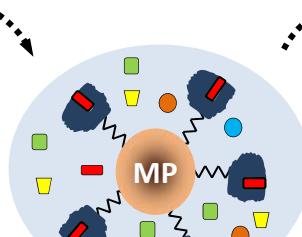
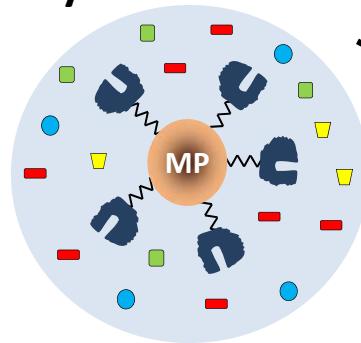
product

inhibitor

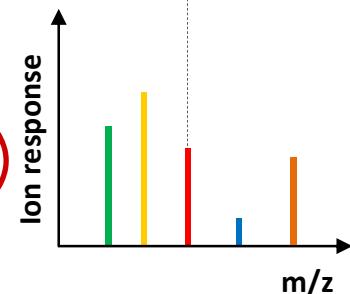
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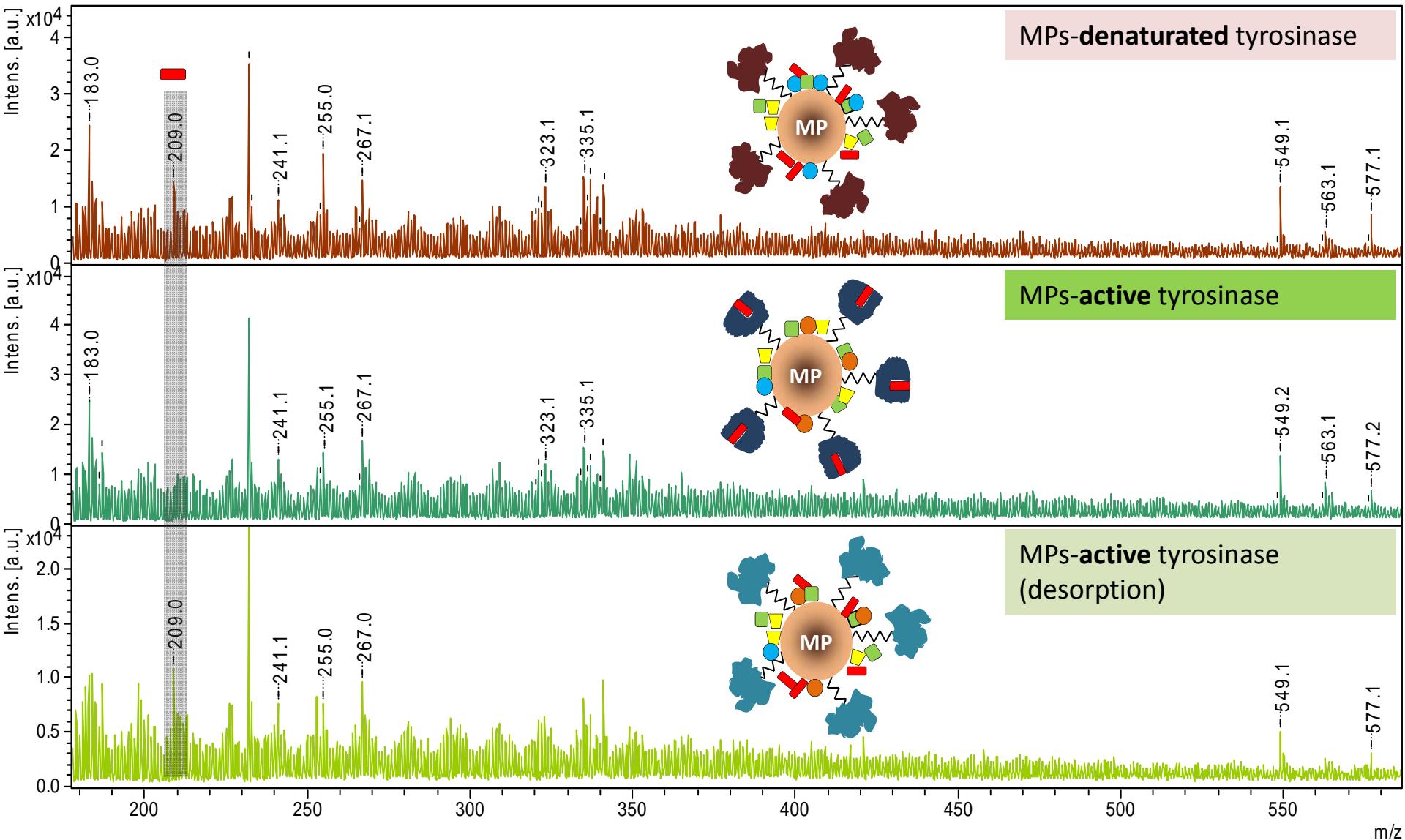
Active enzyme



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Exemplary ENALDI spectra of raw aqueous extract of licorice (*Glycyrrhiza glabra*) root



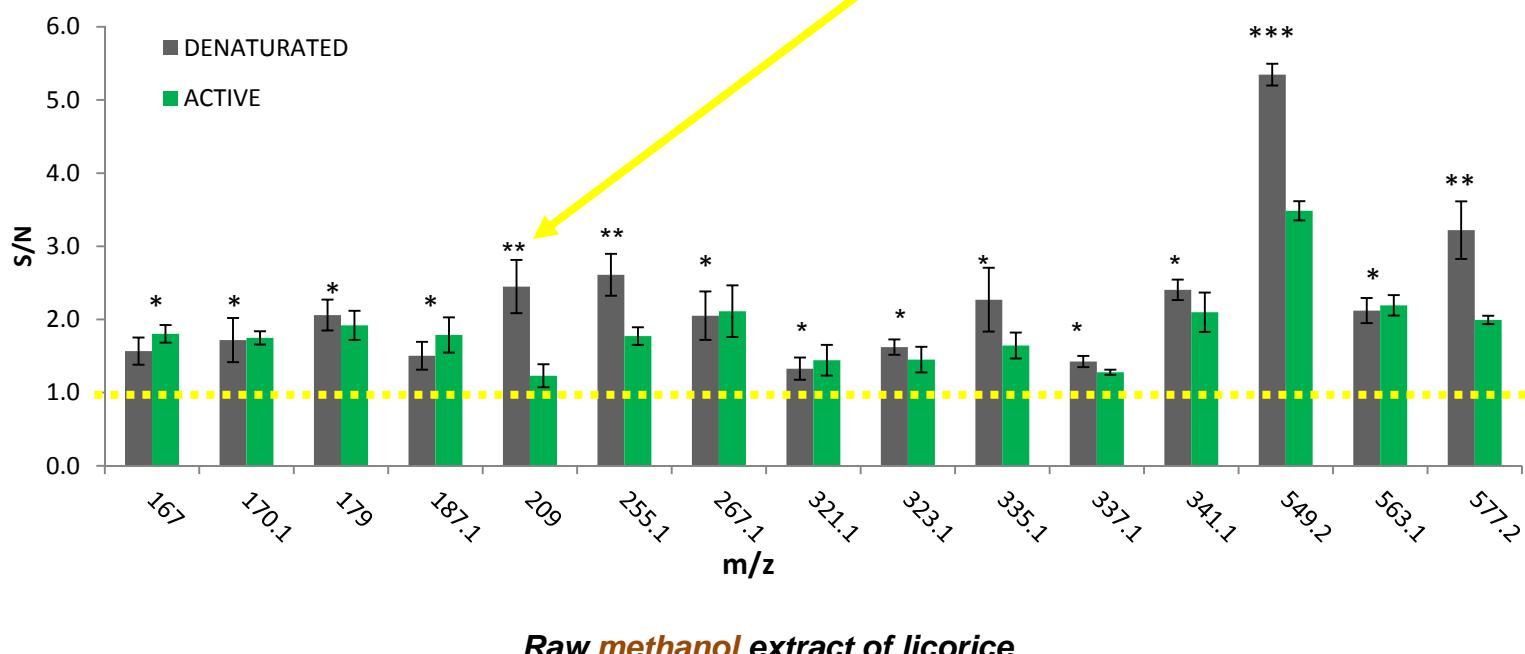
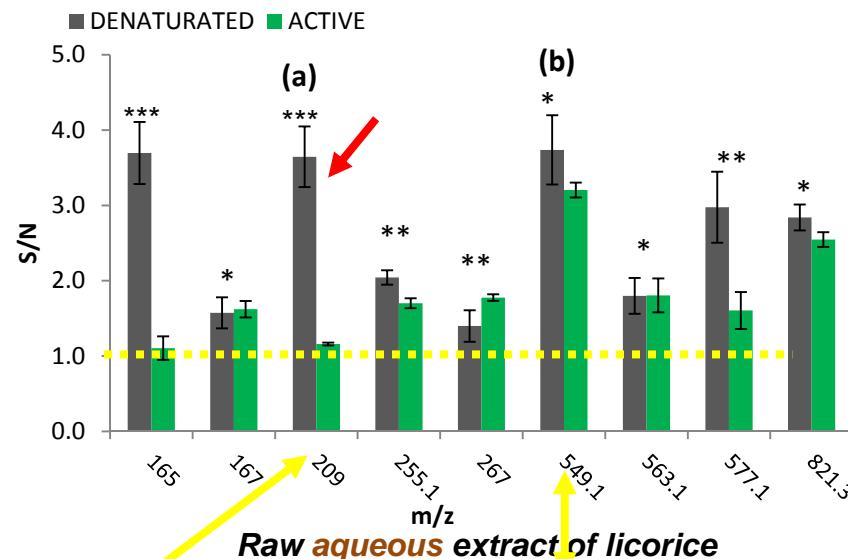
- re-appearance of the ion $[M-H]^-$: 209.0 m/z after treatment of MPs with 0.1% solution of formic acid **directly on the spot**

Statistical description of the data

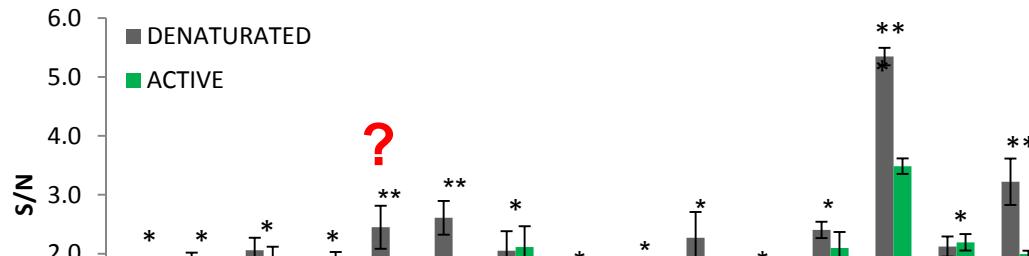
S/N is calculated as the ratio of the intensity of given peak and the adjacent baseline

Student's t-test

- * - $p \geq 5\%$ (S/N not significantly different)
- ** - $p < 5\%$ and $p \geq 1\%$ (S/N significantly different)
- *** - $p < 1\%$ (S/N highly significantly different)



Searching for the inhibitor



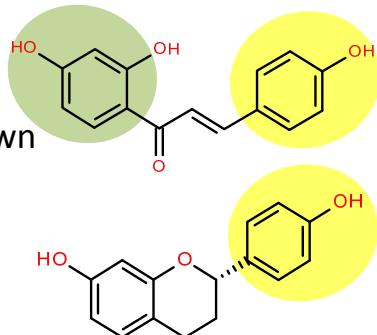
M.A. Farag et al./Phytochemistry 76 (2012) 60–72

Table 2

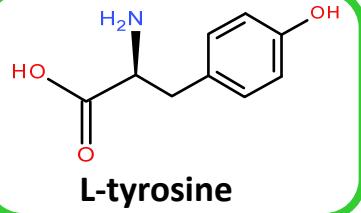
Compounds assigned in *Glycyrrhiza* species methanol extract by LC-MS.

No.	rt (min)	UV max	Identification	Aglycone class	[M-H] ⁻ (m/z)	Error(ppm)	El. Comp.	MS ⁿ ions
1	5.58	275	Unknown		209.0453	+1.14	C ₁₀ H ₉ O ₅	165, 121
2	9.22	270, 315	Glucoliquiritin	Flavanone	579.1698	-1.5	C ₂₇ H ₃₁ O ₁₄	417, 255

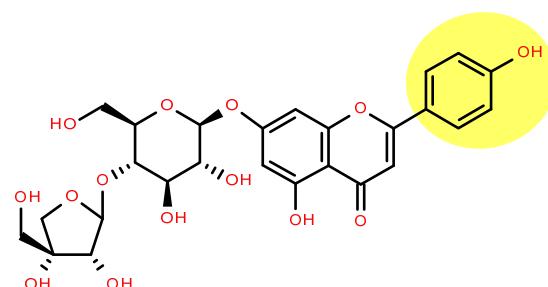
Isoliquiritigenin, known tyrosinase inhibitor^[1]



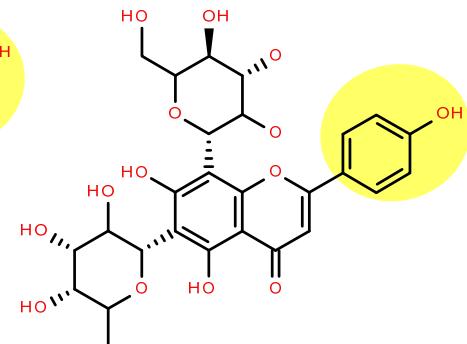
Liquiritigenin



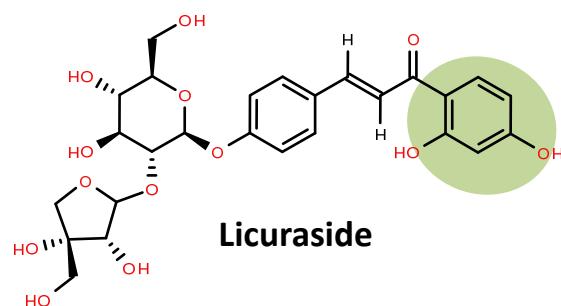
L-tyrosine



Liquiritin apioside

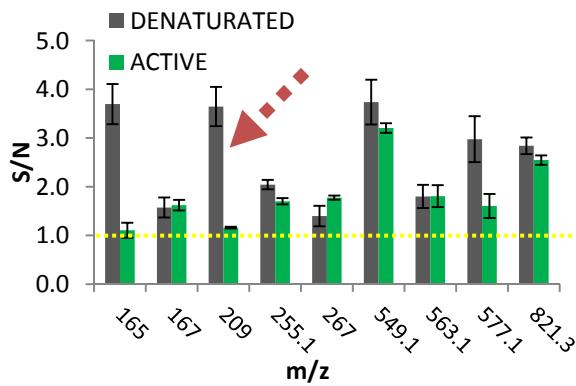


Isoviolanthin

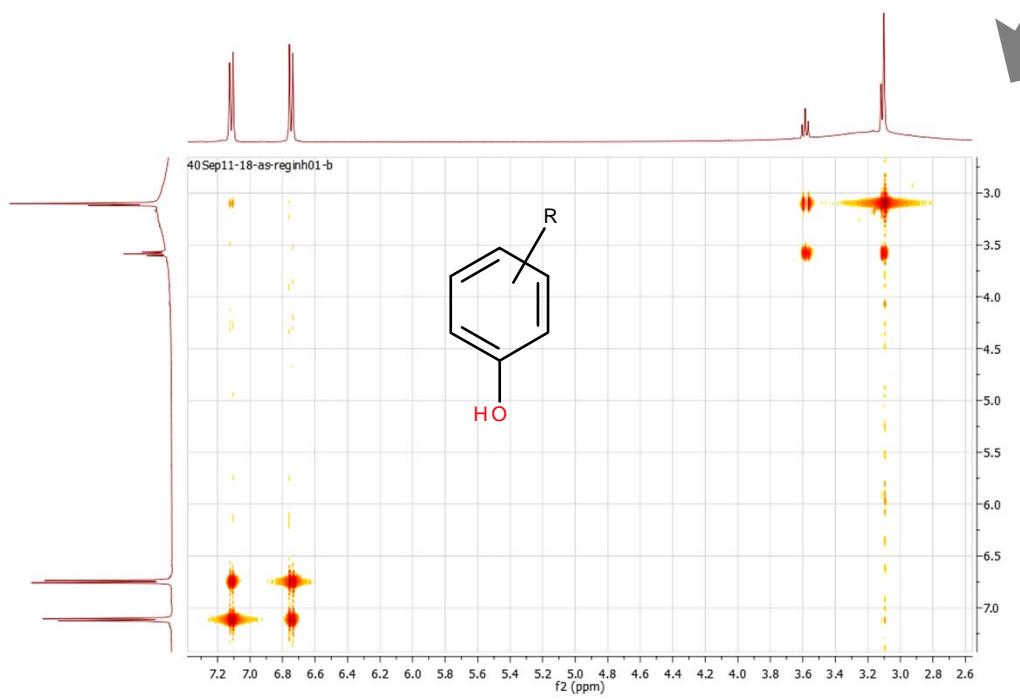
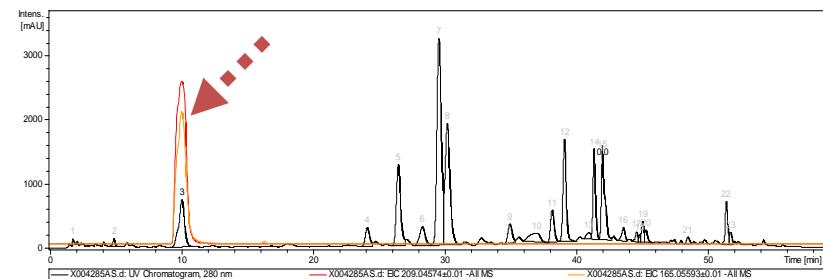


Licuraside

Structural identification of the inhibitor

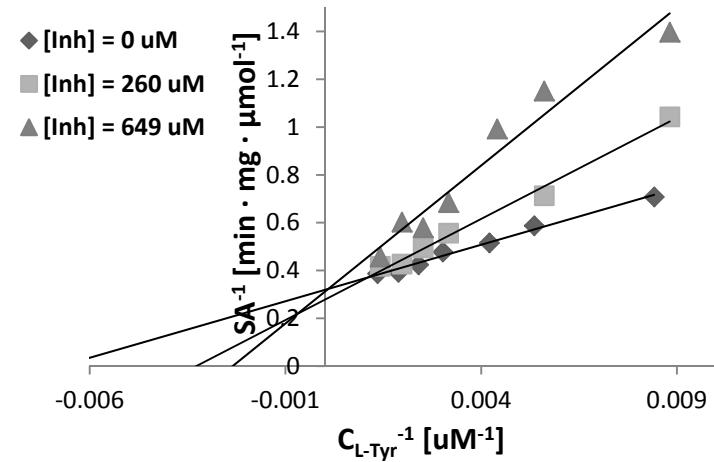
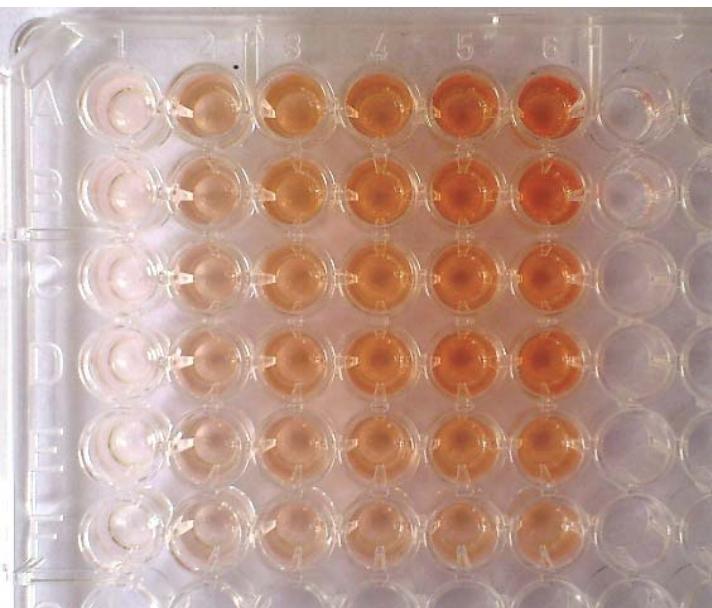


PE separation in LC/MS mode;
localizing the inhibitor candidate; purification



Structure identification by NMR study (¹H, ¹³C, HSQC, COSY)

Kinetic test of purified compound



$$K_I = 291 \mu\text{M}$$

Conclusions

Advantages of ENALDI approach:

- simplified study of complex extracts (no pre-treatment, purification is limited to prospective inhibitors, no laborious fractionation required);
- high stability of MPs (45% of initial activity after 80 days of storage in 4°C);
- high homogeneity of the sample: no ‘sweet spots’ on the MALDI plate;
- possibility to distinguish substrate and inhibitor;
- magnetic properties of beads: easy purification, no centrifuge required;
- low protein consumption per analysis (approx. 6.5 pmol)

Further development

- improvement of S/N ratio in low mass range